The assessment of facial variation in 4747 British school children

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SUMMARY The aim of this study is to identify key components contributing to facial variation in a large population-based sample of 15.5-year-old children (2514 females and 2233 males).

The subjects were recruited from the Avon Longitudinal Study of Parents and Children. Threedimensional facial images were obtained for each subject using two high-resolution Konica Minolta laser scanners. Twenty-one reproducible facial landmarks were identified and their coordinates were recorded. The facial images were registered using Procrustes analysis. Principal component analysis was then employed to identify independent groups of correlated coordinates.

For the total data set, 14 principal components (PCs) were identified which explained 82 per cent of the total variance, with the first three components accounting for 46 per cent of the variance. Similar results were obtained for males and females separately with only subtle gender differences in some PCs.

Facial features may be treated as a multidimensional statistical continuum with respect to the PCs. The first three PCs characterize the face in terms of height, width, and prominence of the nose. The derived PCs may be useful to identify and classify faces according to a scale of normality.

Introduction

Facial morphology attracts interest from a wide variety of research disciplines (e.g. anthropology, developmental anatomy, genetics, maxillofacial surgery, cosmetic surgery, orthodontics, and psychology). The three-dimensional (3D) analysis of facial soft tissues has potential for the identification of morphological features and changes in these features as a result of growth and clinical interventions (McCance *et al.*, 1992; Ferrario *et al.*, 1998a,b, 1999; Nute and Moss, 2000; Hennessy and Moss, 2001; Moss *et al.*, 2003; Kau and Richmond, 2008).

Recent research has focused on facial landmark variation particularly in genetic and medical disorders (Hennessy et al., 2002, 2004, 2007, 2010; Hammond et al., 2004, 2005). There are many theories of facial growth and irrespective of the theory, development of the face can be influenced by advantageous or adverse events particularly during the gestation period extending into early childhood. The genetic and environmental interactions may result in minor physical anomalies affecting the relative position of facial landmarks.

However, the differences in the anatomical position of facial landmarks are usually very subtle and difficult to recognize by subjective examination alone. There are a number of algorithms available that evaluate 3D facial landmarks, which are commonly affected in craniofacial syndromes (Shaner *et al.*, 2000; Bugaighis *et al.*, 2010). The relative position of facial landmarks has been effectively

used to identify some genetic disorders including Williams, Smith-Magenis, 22q11 deletion, and Noonan syndromes (Hammond *et al.*, 2004, 2005).

Although there has been significant attention to 3D assessment of discrete craniofacial anomalies, less attention has been paid to assess normal variation in a population.

The aim of the study is to identify key components contributing to facial variation in a large population-based sample of 15.5-year-old children (2514 females and 2233 males).

Subjects and methods

The children involved in this study were recruited from the Avon Longitudinal Study of Parents and Children (ALSPAC), which was designed to explore how the individual's genotype interacts with different environmental factors to influence health, behaviour, and development of children (Golding *et al.*, 2001). The initial ALSPAC sample consisted of 14541 pregnancies. This was the number of pregnant women enrolled in the ALSPAC study with an estimated date of delivery between April 1991 and December 1992. Out of the initial 14541 pregnancies, all but 69 had known birth outcome. Of these 14472 pregnancies, 195 were twins, 3 were triplets, and 1 was a quadruplet pregnancy; meaning that there were 14676 fetuses in the initial ALSPAC sample. Of these 14676

fetuses, 14062 were live births and 13988 were alive at 1 year.

The cohort was recalled when the children aged 15.5 years. Invitations were sent to 9985 participants who reported that they were interested to take part in the clinics.

Ethical approval for this study was obtained from the ALSPAC Law and Ethics Committee and the Local Research Ethics Committees.

3D facial scans of the children were taken using two high-resolution Konica Minolta VI-900 laser scanners, 'Konica Minolta Sensing Europe, Milton Keynes, UK'. The cameras were fitted with lenses of focal length 14.5 mm and were connected in serial via a Small Computer System Interface cable to a desktop computer workstation. The scanning of each participant took approximately 8 seconds. The set of left and right facial scans of each individual were processed, registered, and merged using a locally developed algorithm implemented as a macro in 'Rapidform® software; INUS Technology Inc., Seoul, South Korea' (Kau et al., 2004; Zhurov et al., 2005; Toma et al., 2008; Kau and Richmond, 2010).

Twenty-one soft tissue landmarks as defined by Farkas (1994) were manually identified on each facial shell using Rapidform® software (Figure 1). The x, y, and z coordinates of each landmark were recorded (63 coordinates in total). The reproducibility of these landmarks in the x, y, and z dimensions has been previously investigated and the majority showed an error of less than 1 mm for both intraand inter-examiner assessments (Toma $et\ al.$, 2009).

Statistical analysis

All facial shells were initially normalized to natural head posture with the origin set at mid-endocanthion point (Figure 2), the *x*-axis pointing left, from right to left eye;

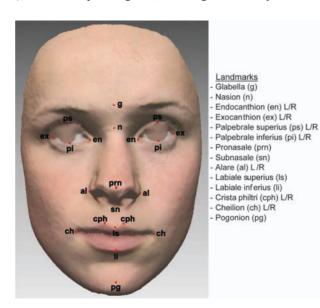
y-axis pointing vertically upwards, from chin to forehead; z-axis pointing outwards, in the nose direction; and the coronal, sagittal, and transverse planes were taken as the xy, yz, and xz planes, respectively (Toma et al., 2009; Zhurov et al., 2010).

Generalized Procrustes analysis was performed to register (align) the sets of 21 facial landmarks by removing translation and rotation (Bookstein, 1991). Scaling was not performed to preserve face size. A centroid representing the mean position for each of the 21 clusters of landmarks was derived. In total, there were 21 centroids. The standard deviations around each centroid were calculated for all individuals and plotted as ellipsoids (Figure 3). Each ellipsoid covers 2 SDs from the mean in x, y, and z, i.e. representing 95 per cent of the variability.

Principal component analysis (PCA) was then employed to identify relatively important correlated variables (Pearson, 1901; Hammond *et al.*, 2004, 2005). The 'Kaiser–Guttman criterion' (Guttman, 1954; Cliff, 1988; Jackson, 1993) was used as the stopping rule to identify critical principal components (PCs). According to this rule, the PCs with eigenvalues greater than the average eigenvalue should only be retained. The rotation method used for PCA was varimax with Kaiser normalization (Kaiser, 1958).

Visualizing facial variation

Average faces were constructed using a locally developed algorithm (Zhurov *et al.*, 2010) implemented as a Rapidform® macro. Parameters characterizing the first PCs, which describe the majority of facial variation, were derived (see the end of the Results) and all faces were split into seven groups in each of the parameters, corresponding to -3 through +3 SDs from the mean value. All facial shells were



Definition Most prominent midline point between eyebrows Deepest point of nasal bridge Inner commissure of the left and right eye fissure Outer commissure of the left and right eye fissure Outer commissure of the left and right eye fissure Superior mid-portion of the free margin of upper left and right eyelids Inferior mid-portion of the free margin of lower left and right eyelids Most protruded point of the paex nasi Mid-point of angle at columella base Most lateral point on left and right alar contour Mid-point of the upper vermilion line Mid-point of the lower vermilion line Point on left and right lebvated margins of thephiltrum just above VL Point located at left and right labial commissure Most anterior mid-point of the chin

Figure 1 Facial soft tissue landmarks.

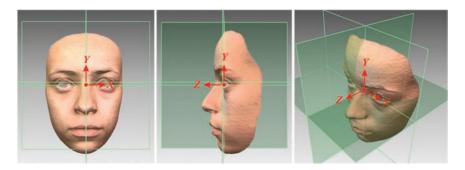


Figure 2 Normalization of facial shells to natural head posture; the *x*-axis (horizontal); *y*-axis (vertical); *z*-axis (depth of field); the coronal, sagittal, and transverse planes were taken as the xy, yz, and xz planes, respectively.

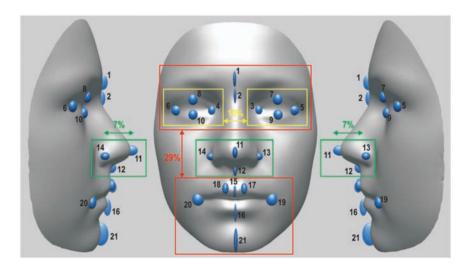


Figure 3 Standard deviation envelopes for 21 facial landmarks, highlighting facial morphology variation revealed by the first three principal components (PCs): PC1 (red, explains 29% of total variance), PC2 (yellow, 10%), and PC3 (green, 7%). 'Facial landmarks': 1, glabella; 2, nasion; 3 and 4, endocanthion (left and right); 5 and 6, exocanthion (left and right); 7 and 8, palpebrale superius (left and right); 9 and 10, palpebrale inferius (left and right); 11, pronasale; 12, subnasale; 13 and 14, alare (left and right); 15, labiale superius; 16, labiale inferius; 17 and 18, crista philtri (left and right); 19 and 20, cheilion (left and right); 21, pogonion.

aligned so that their mid-endocanthion points coincide as well as the sagittal, coronal, and transverse planes.

Results

A total of 5253 children attended the recall clinic. There were 506 individuals out of this sample, which were excluded due to not having their facial images recorded at the time of attending the clinic, poor quality facial scans, obvious ethnicity, and obvious facial dysmorphology. The sample represented normal variation in 4747 Caucasian children (2514 females and 2233 males). The ellipsoid envelopes of variation for the 21 facial landmarks are displayed in Figure 3. The pogonion showed the largest variation in the *y* and *z* axes, whereas the inner canthi and the left and right alari exhibited the least variation.

For the total sample, 14 PCs were identified by PCA (Table 1). This table lists the factor loadings (coefficients) for each coordinate in all extracted PCs. These coefficients indicate the relative importance of different landmark coordinates to the variation identified by each component. Each component includes a group of landmark coordinates (highlighted cells) that have high loadings (coefficients >0.5 in magnitude) in the rotated component matrix. These landmark coordinates contribute greatly to the variation identified by each component. The non-highlighted cells within each component (coefficients < 0.5 in magnitude) indicate landmark coordinates that have less effect on facial variation (coefficients in the range 0.1–0.49 in magnitude are presented and coefficients < 0.1 in magnitude are not shown).

The 14 PCs explain 82.1 per cent of the total variance in facial form (Table 2), with the first three components

 Table 1
 Principal component analysis of landmark data—4747 individuals. PC, principal component

С	X-Y-Z	PCs													
		1	2	3	4	5	6	7	8	9	10	11	12	13	14
C1	lsY	-0.851		0.260			0.102				0.186				
	cphRY	-0.843 -0.841		0.243 0.240			0.102 0.107				0.126 0.137				
	cphLY enLY	0.834	0.138	-0.112	0.165	-0.200	-0.155				0.137				-0.10
	enRY	0.829	0.160	-0.108	0.153	-0.176	-0.180						0.106		0.10
	pgY	-0.822	-0.155	*****		*****	-0.187								
	chRY	-0.816	-0.190		-0.214										
	chLY	-0.814	-0.180		-0.221			-0.100						-0.121	
	piRY	0.810			0.154	-0.151	-0.256						0.125	-0.100	0.16
	piLY	0.808			0.165	-0.170	-0.254						-0.137	-0.104	-0.16
	psLY	0.792					-0.278						-0.123	0.332	-0.12
	psRY liY	0.783	0.124		0.296		-0.276						0.112	0.335	0.18
	exRY	-0.769 0.759	-0.124		-0.386 0.189	-0.210	-0.309						0.145		0.19
	exLY	0.748			0.194	-0.173	-0.313						-0.184		-0.22
	gY	0.644	0.223		0.174	0.308	-0.130					-0.192	0.104		0.22
	nY	0.620	0.132	-0.468		0.123	0.150					-0.145			
C2	psLX	0.143	0.939						-0.125						
	psRX	-0.139	-0.939						-0.116						
	piLX	0.144	0.933						-0.135						
	piRX	-0.150	-0.932						-0.127				-0.126		
	enRX		-0.837			0.129			-0.168					0.219	
	enLX	0.102	0.830			-0.108		0.122	-0.126			0.117		-0.225	
	exRX exLX	-0.192 0.184	-0.810 0.768					0.123 -0.123	-0.148			-0.116 0.140		-0.391 0.432	
C3	alLZ	0.184	0.700	-0.798		-0.101	-0.171	-0.123 -0.130	-0.148			-0.136		0.432	
-3	alRZ	0.220		-0.786		0.101	-0.152	-0.144				-0.130			
	snZ	0.347		-0.706	0.260	-0.144	0.102	-0.150				-0.113			
	prnZ	0.404		-0.690	0.200	-0.253	-0.138	-0.204				-0.185			
	liΖ	0.295		0.562				-0.239							
C4	lsZ	0.368			0.863			-0.102			-0.163				
	cphRZ	0.391			0.861	-0.112		-0.110							
	cphLZ	0.388			0.861	-0.117							-0.109		
a.	pgZ	-0.151	0.100	0.271	-0.788	0.050	-0.195	-0.321				0.140		0.150	
C5	gZ	-0.103	0.198	0.176		-0.858	0.102					-0.148		0.158	
	nZ piRZ	-0.374	0.157	-0.163 0.352	-0.245	-0.822 0.673	0.103				0.114	-0.299 -0.120		0.167	
	piLZ	-0.374		0.332	-0.243 -0.242	0.659			0.117		0.114	-0.120 -0.111	0.119		
	enLZ	-0.469		0.244	-0.191	0.521	0.154		0.111		0.105	-0.139	0.117	0.274	
	enRZ	-0.458		0.250	-0.185	0.485	0.167		*****	-0.105		-0.140		0.321	
C6	prnY					0.120	0.821					0.106			
	alLY	-0.284					0.791		-0.108						
	alRY	-0.299					0.768								
	snY	-0.226	0.120	0.184			0.722	0.02#			0.106	0.162			
C7	chRX	-0.123	-0.128					0.835			0.196				
	chLX chLZ	0.139 0.203	0.145	0.230				-0.821 0.814			-0.205		-0.124		
	chRZ	0.203		0.230				0.814					0.124		
C8	snX	0.173		0.223				0.000	0.940				0.172		
	prnX								0.906	0.167					-0.13
C9	gX									0.974					
	nX									0.967					
C10	cphRX	-0.164						0.183			0.809		0.312		
	cphLX	0.147						-0.199			-0.768		0.422		
	alLX	0.266	0.321		0.158	-0.163		-0.219	0.266		-0.488	0.188			
	alRX	-0.275	-0.332		-0.162	0.158		0.219	0.265		0.481	-0.189			
C11	psLZ	-0.275		0.207		0.101	0.219		0.109			0.805	0.109		
212	psRZ	-0.288		0.237			0.209		-0.116			0.784	-0.125		
C12 C13	lsX exLZ	-0.307	-0.252	0.278	-0.172	0.369			0.166	0.105			0.942 0.161	-0.618	
C13	exLZ exRZ	-0.307 -0.320	-0.252 -0.290	0.278	-0.172 -0.186	0.369	0.109		-0.129	-0.118			-0.146	-0.598	
C14	pgX	0.520	0.270	0.207	0.100	0.500	0.107		-0.129	0.110			-0.153	0.070	0.92
	liX								-0.104				0.486		0.77

The highlighted cells (coefficients >0.5 in magnitude) indicate landmark coordinates that contribute greatly to the facial variation; non-highlighted cells (coefficients <0.5 in magnitude) indicate landmark coordinates that have less effect on facial variation (coefficients in the range 0.1–0.49 in magnitude are presented and coefficients <0.1 in magnitude are not shown).

Table 2 Brief description of the principal components (PCs) extracted for the total sample and their corresponding positions in male and female samples

Total sample $(N = 4747)$	Males $(N = 2233)$		Females $(N = 2514)$		
Brief description of PCs	Percentage of variance	PC	Percentage of variance	PC	Percentage of variance
PC1, face height	28.8	PC1	24.2	PC1	21.9
PC2, inter-eye distance (face width)	10.4	PC2	11.0	PC2	11.1
PC3, prominence of the nose	6.7	PC3	7.4	PC3	7.6
PC4, protrusion of the upper lip relative to the chin	5.3	PC4	5.4	PC4	5.7
PC5, eye depth relative to the nasal bridge	4.8	PC6	4.5	PCs 7 and 9	4.4 + 3.3
PC6, vertical height of the nose	4.4	PC5	5.1	PC6	4.4
PC7, ratio of the mouth width to mouth depth	4.0	PC7	4.4	PC5	5.0
PC8, deviation of the nasal tip and columella base	3.6	PC8	3.8	PC8	3.9
PC9, horizontal asymmetry of the nasal bridge	3.2	PC10	2.7	PC10	2.9
PC10, philtrum-to-nose width ratio	2.7	PCs 14 and 15	1.8 + 1.7	PCs 15 and 16	1.7 + 1.6
PC11, Upper eyelid depth	2.4	PC12	2.3	PC13	2.0
PC12, horizontal asymmetry of the upper lip (philtrum)	2.3	PC11	2.5	PC11	2.7
PC13, Facial flatness (outer canthi depth)	1.9	PC9	3.4	PC14	1.8
PC14, horizontal asymmetry of the chin and lower lip	1.7	PC13	2.0	PC12	2.4

The order of PCs (1–14) for the total sample is based on their percentage of variance (descending order), PC1 has the highest percentage of variance and PC14 has the least percentage of variance. PCs 14 and 15 (males) describe variation in philtrum and nose width, respectively; PCs 7 and 9 (females) describe variation in depth of the lower eyelids (relative to the nasal bridge) and inner canthi, respectively; PCs 15 and 16 (females) describe variation in nose and philtrum width, respectively. Examples: PC6 (total sample) explains variation of the vertical height of the nose is positioned as PC5 (males) and PC6 (females); PC7 (total sample) explains variation of the mouth width to mouth depth ratio is positioned as PC7 (males) and PC5 (females).

accounting for 45.9 per cent of the total variance (PC1 28.8 per cent, PC2 10.4 per cent, and PC3 6.7 per cent). The other PCs account for considerably smaller portions of the total variance (PC4 5.3 per cent, PC5 4.8 per cent, PC6 4.4 per cent, etc.).

The first PC includes two subsets of landmarks grouped around the eyes and mouth (highlighted by red rectangles in Figure 3). The first subset represents the *y* coordinates of 10 upper face landmarks including eight landmarks around the eyes (3–10) as well as glabella and nasion. The second subset includes the *y* coordinates of seven lower face landmarks (15–21). The loadings of the two subsets have opposite signs; this indicates statistical variation in opposite (upward–downward) directions. Therefore, PC1 essentially describes variation in face height.

The second PC (enclosed in yellow rectangles) consists of the *x* coordinates of eight landmarks around the eyes (3–10). Loadings with opposite signs correspond to variation in opposite (outward–inward) directions. Therefore, this component essentially describes variation in inter-eye width.

The third PC (indicated by green rectangle) represents a single group of the z coordinates of four landmarks associated with the nose (11–14); consequently, this component characterizes the prominence of the nose.

PCA was also applied when the data set was separated out into males and females. Fifteen PCs were identified for males and 16 for females. Brief component definitions and variances explained are listed in Table 2. The first eight PCs for males and the first four components for females were roughly the same as those of the total sample. Subtle gender

differences were noticed in the sequence of some PCs as compared with the total sample; for example, PC14 (related to asymmetry of the chin in the total sample) was positioned as PC13 and PC12 in males and females, respectively.

Visualizing facial variation

A total of 21 average faces were constructed to visualize facial morphology variation identified by the first three PCs (Figure 4). Three parameters (P1, P2, and P3) characterizing the first three PCs were defined as follows:

- P1: vertical distance between the centroids of the upper and lower sets of landmarks (1–10 and 15–21);
- P2: horizontal distance between the centroids of the left and right sets of landmarks associated with the eyes;
- P3: *z* coordinate of the centroid of the landmarks associated with the nose (11–14).

The facial variation identified by the first three PCs is presented visually in this section as these components explained the majority of the variation and it is shown as an example that the principle can be applied to all 14 components.

Discussion

Fifty-three per cent of the invited children attended the recall, and 90 per cent of these had suitable facial scans. The present study assessed normal variation of facial morphology in a large population-based cohort of Caucasian adolescents 15.5 years of age. The results can be considered specific to

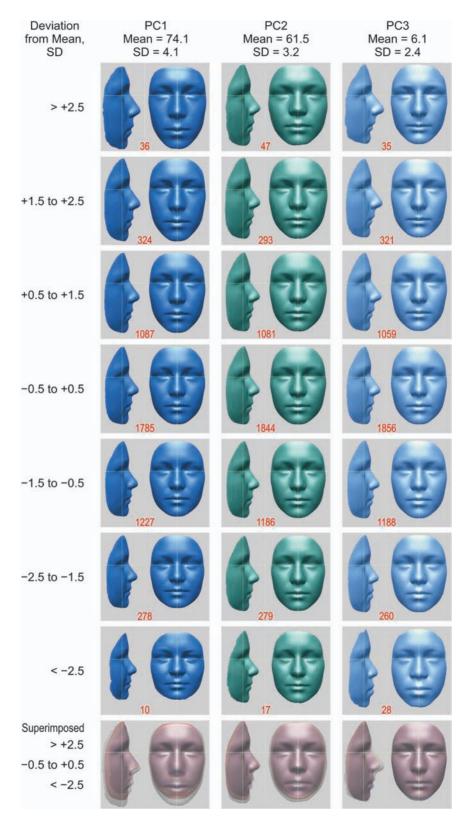


Figure 4 Average faces constructed to illustrate variation in face height (left column, PC1), intereye distance (middle column, PC2), and prominence of the nose (right column, PC3). The numbers shown in red colour represent the number of individuals contributed to each average face.

this particular population, and the methodology used in this study can form the basis to analyse and compare facial morphology of other population groups.

The 3D acquisition technique used in this study is laser scanning; the accuracy of surface scanning with Konica Minolta 900/910 laser scanners is in the range of 0.3–0.5 mm, whereas the images obtained with a photogrammetry approach such as 3dMD (Atlanta, GA) cameras have been reported as 0.8–1.0 mm (Kau *et al.*, 2005). The laser scanner has sufficient surface resolution to detect the detailed morphology, particularly the fine lines that form the inner and outer canthi. However, the capture time of the 3dMD system is 1.5 milliseconds at the highest resolution compared to approximately 8 seconds scanning time using laser scanners. Therefore, the laser scanning requires a protocol to instruct the patient to remain still, presenting with no facial expressions.

Although the acquisition time for the laser cameras is relatively long, the reliability of this technique is remarkably good over a 3 day period (Kau *et al.*, 2005). Both Konica/Minolta 900/910 and the photogrammetry techniques use colour texture overlays, and the resolution of the colour texture is superior using photogrammetry compared with the laser technique. However, achieving consistent colour balance is difficult and often the colour tones can mask facial contours and in this study the colour texture was not applied.

Fourteen PCs were identified describing the majority of facial soft tissue variation (82 per cent), with the first three components accounting for 46 per cent of the total variance. The sample was registered using Procrustes analysis; with this technique, the landmark 3D coordinates were placed in the same space reducing confounding errors (rotation and translation).

There have been several studies using PCA on either lateral skull radiographs or photographs. One of these studies assessed craniofacial form in 622 subjects and identified six PCs that explained 68 per cent of the variation. The study did not use Procrustes analysis to register the landmarks and arguably resulted in a rather complex array of facial parameters forming each PC (Cleall *et al.*, 1979). However, the first and third PCs were broadly similar to the findings in the present study, the first representing face height and the third convexity (mid-face and dental protrusion). The second component related to anteroposterior aspects of facial morphology, which is recorded in PC4 in the present study.

Photographs were used to identify six components explaining 86.5 per cent of the variance (Krey and Dannhauer, 2008). The first PC (33.9 per cent) described scaling along an axis from Porion to chin (a combination of vertical and horizontal vectors); the second component (28.6 per cent) characterized the vertical dimension of the lower face.

The soft tissue profiles of 170 patients aged 7–17 years were assessed (Halazonetis, 2007). The first eight PCs

explained 90 per cent of the total shape variability. The first component (36 per cent) related to lip, nose, and chin prominence; the second component (18 per cent) related to facial convexity; and the next two components mainly related to lower lip shape. The overall shape differences between the average profiles of boys and girls were minor.

There were similarities and differences identified when making a comparison with previous studies. The present study was undertaken on a large cohort of the same age and previous studies have included subjects ranging from 7 years of age to adulthood. In addition, previous studies have used 2D records, whereas the 3D data utilized in this study should eliminate projection problems commonly found in radiographs and photographs (Houston *et al.*, 1986; Benson and Richmond, 1997). The chin prominence reported by Halazonetis (2007) and Krey and Dannhauer (2008) would be reported as a positive change in the *z*-axis for upper lip landmarks relative to the chin in PC4 of the current study, although this component only explains 5.3 per cent of the total variance.

As the 3D data were registered in a common space using Procrustes analysis, the derived PCs should be more valid based on relative importance of independent landmark coordinates in space. The 14 PCs derived from the current study reflect the complexity of facial morphology. The first three components describing face height, width, and convexity with the other 11 PCs contribute subtle changes to the face that makes the face unique.

The first PC (face height) explained 29 per cent of the variance and this evidence gives support to previous facial classifications as long/thin and short/wide faces (Schendel et al., 1976; Opdebeeck and Bell, 1978; Opdebeeck et al., 1978; Farkas, 1994). In this study, the average distance between the upper and the lower facial centroids (parameter P1) was 74.1 mm (ranging from 59.8 to 91.6 mm) with the nasion to pogonion distance at 101.7 mm (ranging from 82.8 to 127.6 mm). Male faces were on average 6 mm longer than female faces. This distance is slightly less than 8 mm reported for 50 fifteen-year-old Caucasians assessed by Farkas (1994) and higher than 1.8 mm for approximately 40 norms, 8- to 12-year old (Bugaighis et al., 2011). In addition, previous clinical studies of long and short face types also reported limited samples, which reflect face heights equivalent to 2 SDs from the mean (Schendel et al., 1976; Opdebeeck and Bell, 1978).

For PC2, the average distance between left and right centroids of the landmarks associated with the left and right eyes (parameter P2) was 61.5 mm (range 50.5–73 mm) with the average distance between the inner canthi of the eyes 34.2 mm (range 24.0–46.5 mm). The inter-canthal distance on average was 1.2 mm larger in males compared to females. Similar findings have been reported in smaller samples (Laestadius *et al.*, 1969; Farkas, 1994; Bugaighis *et al.*, 2011). Many syndromes exhibit abnormal inter-eye distance that may even exceed 2 SD from the mean (Feingold and

Bossert, 1974; Farkas *et al.*, 1989; Cohen *et al.*, 1995; Miamoto *et al.*, 2011); hypertelorism can be seen in 1q21.1 duplication syndrome, apert syndrome, basal cell nevus syndrome, Crouzon syndrome, DiGeorge syndrome, Noonan syndrome, and LEOPARD syndrome (Mann, 1957; Kreiborg and Cohen, 2010; Randolph *et al.*, 2011); and hypotelorism can be seen in patients with trigonocephaly (Nagasao *et al.*, 2011) and Schilbach–Rott syndrome (Joss *et al.*, 2002).

For PC3, the prominence of nose centroid (parameter P3) was on average 6.1 mm (ranging from -5.8 to 17.3 mm). The nasal tip protrusion (subnasale–pronasale) was on average slightly less in females (19.4 mm) compared to males (20.1 mm). Similar findings were reported elsewhere (Farkas, 1994; Zankl *et al.*, 2002).

Parameters P1, P2, and P3 associated with the first three PCs can be used to characterize the face as a 3D statistical continuum, where each coordinate corresponds to the standard deviation from the mean value of the respective parameter. Although male faces are generally larger than female faces (Ferrario *et al.*, 1998a,b, 1999), the PCAs for males and females show similar relative importance of facial parameters, which will be useful in facial classification.

Facial asymmetry has been suggested to arise from random variation or genetic and environmental influences (Waddington, 1957). The present study showed that minor facial asymmetry is relatively common in both sexes with similar patterns (PC8, nasal tip/columella base; PC9, nasal bridge; PC12, upper lip/philtrum; PC14, lower lip/chin). A mild degree of facial asymmetry has been reported elsewhere (Lu, 1965; Vig and Hewitt, 1975; Shah and Joshi, 1978; Alavi *et al.*, 1988; Peck *et al.*, 1991; Pirttiniemi, 1992; Ferrario *et al.*, 1993). Differences in facial asymmetry have been reported between the sexes; however, most of these studies have been undertaken on small samples (Farkas and Cheung, 1981; Ferrario *et al.*, 1994, 2001; Severt and Proffit, 1997; Shaner *et al.*, 2000; Smith, 2000; Haraguchi *et al.*, 2002; Hardie *et al.*, 2005; Ercan *et al.*, 2008).

In this study, the chin point (pogonion) deviated between –5.6 and 5.2 mm from the sagittal plane; nasal tip (pronasale), –4.7 and 4.9 mm, compared to the columella base (subnasale), –3.1 and 2.6 mm; glabella, –2.9 to 2.0 mm; nasion, –2.2 to 1.9 mm; upper lip (labiale superius), –2.3 and 2.5 mm; lower lip (labiale inferius), –1.5 to 2.8 mm. In previous studies, the degree of asymmetry has been attributed to discernible imbalances in the development of skeletal, dental, and soft tissues (Williamson and Simmons, 1979; Alavi *et al.*, 1988; Pirttiniemi *et al.*, 1990; Schmid *et al.*, 1991; Pirttiniemi, 1992). Unfortunately, the methods employed in these studies describe details of local imbalances with less emphasis on systematic assessment of facial asymmetry.

The current study provides a comprehensive range of soft tissue facial parameters for a population of 15.5-year-old Caucasians. The levels of deviation from the mean for the various parameters provide a basis for future assessment of subjects using craniofacial landmarks. Moreover, facial height and width have been reported to show strong genetic components (Savoye *et al.*, 1998; Baydas *et al.*, 2007) and it is intended to use the current data set in the future to explore genotype/phenotype associations through a genome-wide association study.

There are many projects underway around the world such as the FaceBase Consortium (Hochheiser *et al.*, 2011) collecting both 3D facial images and genetic data with the intention to undertake genome-wide association studies. It is important that the face data collected are standardized with matching age groups to allow analyses within and across population groups.

Conclusions

Fourteen PCs were identified for the total sample, which explained 82 per cent of the total variance in facial form, with the first three components accounting for 46 per cent of the variance. Similar results were identified for the data set when split into males and females, suggesting that the major components of facial variation do not differ between the genders. Variation in facial form can be accurately quantified and described as a multidimensional statistical continuum. This method of facial assessment may be useful to identify and classify faces and facial changes that occur as a result of growth and inform clinicians of appropriate healthcare interventions for specific facial types.

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